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Heterocyclic Aminopyrrolidine Derivatives as Melatonergic Agents

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Abstract—A series of chiral heterocyclic aminopyrrolidine derivatives was synthesized as novel melatonergic ligands. Binding affinity assays were performed on cloned human MT₁ and MT₂ receptors, stably expressed in NIH3T3 cells. Compound **16** was identified as an orally bioavailable agonist at MT₁ and MT₂ melatonin receptors with low vasoconstrictive activity.
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The neurohormone melatonin (*N*-acetyl-5-methoxytryptamine) (Fig. 1), first isolated from bovine pineal gland extracts in 1958,¹ is present in all mammalian species. It is synthesized and secreted primarily by the pineal gland in a circadian manner that closely follows the daily light/dark cycle.^{2,3} It plays a central role in the regulation of circadian rhythms, the modulation of retinal physiology, and the control of seasonal cycles in vertebrates. Melatonin alleviates jet lag by decreasing sleep latency, improving sleep quality, and reducing fatigue, while accelerating the resynchronization of circadian rhythms associated with transmeridian travel.^{4–7} In addition, melatonin has been shown to have anti-tumor properties and has been implicated in immune system responsiveness.⁸

It has been demonstrated that many of the effects of melatonin are mediated through G-protein coupled receptors expressed primarily in the brain, retina, pituitary, and blood vessels.⁹ Cloning of several G-protein coupled melatonin receptor genes has revealed at least

three melatonin receptor subtypes, two of which are defined as MT₁ and MT₂ and have been found in mammals.¹⁰

As a pharmacological tool and therapeutic entity, melatonin is not ideal because of its short biological half-life (about 19 min in rat), contractile effects on vascular smooth muscle, low aqueous solubility, and poor oral bioavailability. In an attempt to develop novel chemical tools to overcome the liabilities of melatonin, as well as understand melatonin receptor function, we report here the synthesis and biological activity of heterocyclic aminopyrrolidine derivatives as novel melatonergic agents. These derivatives were designed to be melatonergic ligands at melatonin receptors. In addition, they have the potential for improved metabolic stability, increased oral bioavailability, and reduced vasoconstrictive activity due to modification of the aromatic methoxy group and amido side chain.

Initially, pyrrolidine **1** (Fig. 1) was prepared to determine the effect of conformationally-constrained structures on the binding affinity. Evaluation of **1** showed good affinity for MT₁ and MT₂ receptors, $K_i = 3.7$ and 2.6 nM, respectively. Unfortunately, this compound had the same vasoconstrictive activity as melatonin itself in assays conducted with rat caudal arteries. In an effort to

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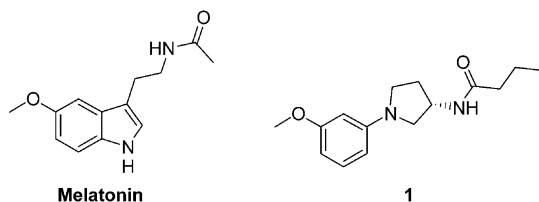
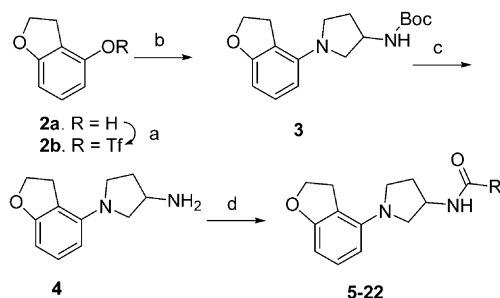


Figure 1.

overcome this liability, the arylalkoxy moiety in **1** was replaced with a dihydrobenzofuran motif. Thus, the targets were redesigned as shown in Scheme 1.¹¹

As depicted in Scheme 1, 4-hydroxydihydrobenzofuran (**2a**) was prepared by Parham cycloalkylation of the 2-bromoresorcinol.¹¹ Phenol **2a** was converted to triflate **2b** by treatment with triflic anhydride in the presence of pyridine. Palladium-catalyzed coupling of triflate **2b** with (*R* or *S*)-pyrrolidin-3-yl-carbamic acid *tert*-butyl ester in the presence of Pd(OAc)₂ and BINAP using Cs₂CO₃ as a base in toluene under reflux furnished carbamate **3**. The yield of the palladium-catalyzed amination step was affected by the percentage of Pd(OAc)₂ and BINAP. When Pd(OAc)₂ (3 mol%) and BINAP (5 mol%) were used, the yield was only 35%. However, when Pd(OAc)₂ and BINAP were increased to 12 mol% and 18 mol%, respectively, the yield was dramatically increased to 86%. Deprotection of **3** with HCl in either ethyl acetate or dioxane gave amine **4** in quantitative yield. The amine **4** was acylated using acid chlorides or reacted with isocyanates to give the desired amide and urea products **5–22**.¹¹

The *K*_i values of compounds **5–22** for human MT₁ and MT₂ melatonin receptor subtypes were determined in binding assays using 2-[¹²⁵I]-iodomelatonin with the described assay method¹² and the results are reported in Table 1. The MT₁ and MT₂ affinity values identified some compounds with excellent affinity but most of the compounds had little selectivity between MT₁ and MT₂ receptors. Cyclopropylcarbamide **9**, in particular, had the highest selectivity for the MT₁ receptor with potent affinity (MT₂/MT₁=9). The data also demonstrated clearly that the affinities of amides **5–14** were related to the absolute stereochemistry: (*S*)-enantiomers were more potent than (*R*)-enantiomers. For example, (*R*)-acetamide **5** had only modest affinity for both MT₁ (*K*_i=202 nM) and MT₂ (*K*_i=121 nM) receptors. How-



Scheme 1. (a) Tf₂O, pyridine, 0–25 °C, 90%; (b) pyrrolidin-3-yl-carbamic acid *tert*-butyl ester, Pd(OAc)₂, BINAP, Cs₂CO₃, toluene, reflux, 86%; (c) HCl/dioxane, 100%; (d) RCOCl, Et₃N, CH₂Cl₂ or RNCO, benzene.

Table 1. Binding affinity of compounds **5–22** for human MT₁ and MT₂ melatonin receptors stably expressed in NIH3T3 cells. Values represent mean from experiments performed in duplicate

Compd	R	Chirality	MT ₁ <i>K</i> _i (nM)	MT ₂ <i>K</i> _i (nM)
Mel	—	—	0.4	0.3
1	—	S	3.7	2.6
5	Me	R	202	121
6	Et	R	49.4	39.3
7	<i>n</i> Pr	R	67	22.3
8	<i>i</i> Pr	R	129	85
9	<i>c</i> Pr	R	3.9	35
10	Me	S	1.5	1.0
11	Et	S	0.18	0.4
12	<i>n</i> Pr	S	0.14	0.31
13	<i>i</i> Pr	S	6.0	6.4
14	<i>c</i> Pr	S	1.2	2.8
15	NH-Me	R	2.2	4.6
16	NH-Et	R	0.65	0.42
17	NH- <i>n</i> Pr	R	4.4	2.6
18	NH- <i>c</i> Pr	R	6.6	10.8
19	NH-Me	S	0.90	0.94
20	NH-Et	S	7.0	2.1
21	NH- <i>n</i> Pr	S	17.7	5.8
22	NH- <i>c</i> Pr	S	21	19

ever, the (*S*)-enantiomer **10** had excellent affinity for both MT₁ and MT₂ receptors (*K*_i=1.5 and 1 nM, respectively). Furthermore, the binding potency also depended on the length or size of the amido chain. Replacement of the acetyl group by propanoyl or butanoyl increased the affinity of the compounds. In contrast to the amide derivatives, the affinity of urea analogues **15–22** was only slightly influenced by the stereochemistry and the length of the alkyl chain, as (*R*)-enantiomers of urea derivatives except (*R*)-urea **15**, which was less active than (*R*)-urea **19**, were slightly more potent than the (*S*)-enantiomers.

The three most active compounds emerging from this series, **11**, **12**, and **16** were evaluated in our advanced profiling assays. It has been reported¹³ that melatonin has a marked ability to enhance α -adrenoceptor-mediated vasoconstriction of the rat tail artery. Thus, the effect of these compounds on vascular smooth muscle was evaluated using the method already described.¹⁴ Compared to melatonin, only compound **16** showed significantly reduced vasoconstrictive activity in assays conducted with rat caudal arteries (0.48 relative to melatonin), whereas **11** and **12** were 1.2 and 0.83, respectively, relative to melatonin. These studies led to the identification of **16** as a potent and promising ligand for MT₁ and MT₂ receptors. Compound **16** was further tested for functional activity in NIH3T3 cells expressing melatonin MT₁ or MT₂ receptor using the method already described¹⁴ and found to be a full agonist at both MT₁ and MT₂ receptors. As Figure 2 shows, compound **16** exhibited dose-dependent inhibition of forskolin-stimulated cAMP accumulation in NIH3T3 cells stably expressing the human MT₁ or MT₂ receptor with a maximum inhibition similar to melatonin.

To determine the selectivity for melatonin receptors over a large number of other hormone or neurotransmitter receptors, enzymes, and ion channels, com-

pound **16** was tested through a commercial screen of radioligand binding panels.¹⁵ At a concentration of 10 μ M, **16** was found to have no significant affinity to all binding sites examined. For these purposes, significant activity was defined as $\geq 50\%$ inhibition of corresponding binding at 10 μ M.

Compound **16** was characterized in several pharmacokinetic studies (Table 2). The oral bioavailability was

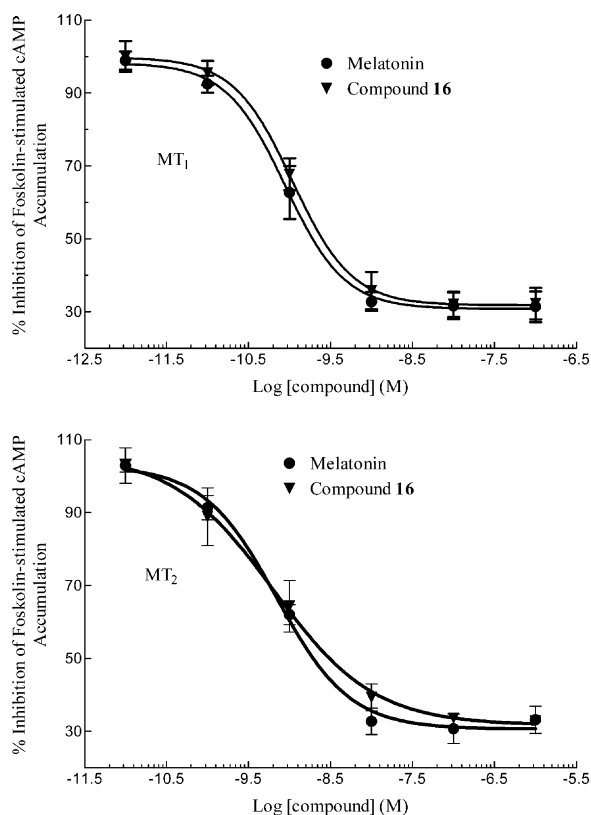


Figure 2. The effect of melatonin and compound **16** on forskolin-induced cAMP accumulation in NIH3T3 cells stably expressing the human MT_1 (upper graph) or the human MT_2 receptor (lower graph). Data is a composite of 3–6 separate experiments performed in duplicate. Non-linear regression analysis yielded for the MT_1 receptor, melatonin $pEC_{50} = 10.1 \pm 0.1$ (0.09 nM), $E_{max} = 69 \pm 2\%$, and intrinsic activity = 1; compound **16** $pEC_{50} = 9.9 \pm 0.1$ (0.13 nM), $E_{max} = 66 \pm 3\%$, and intrinsic activity = 1; for the MT_2 receptor, melatonin $pEC_{50} = 9.1 \pm 0.1$ (0.76 nM), $E_{max} = 69 \pm 2\%$, and intrinsic activity = 1; compound **16** $pEC_{50} = 9.1 \pm 0.24$ (1.1 nM), $E_{max} = 68\% \pm 2\%$, and intrinsic activity = 1.

Table 2. Pharmacokinetic parameters of **16**

PK parameters	Rat ^a	Dog ^b
IV		
Dose (mg/kg)	1	1
$t_{1/2}$ (h)	0.9	1
Cl (mL/min/kg)	15	8.8
Vd (L/kg)	0.7	0.5
PO		
Dose (mg/kg)	1	1
F (%)	49	77

^aCompound dosed in rats as a solution in PEG-400.

^bCompound dosed in male beagle dogs as a solution in 50% PEG-400/50% H_2O .

significant in both rat and dog (49% and 77%, respectively), which is superior to the oral bioavailability of melatonin at the same dose (24% and 17%, respectively). This oral bioavailability is presumably due to its good absorption, consistent with the excellent Caco-2 permeability (P_c 544 nm/s) and moderate clearance. The effective absorption was rapid in the rat, with peak concentration occurring within 30 min, whereas the compound was slowly absorbed in the dog. As a prelude to evaluating compound **16** in animal models of sleep disorder, the ability of compound **16** to enter rat brain following intravenous administration was determined. Table 3 shows the plasma and brain concentrations, as well as brain/plasma ratios at 20 min and 2 h after administration of an IV bolus dose (5 mg/kg). Compound **16** was shown to have moderate brain penetration based on the samples taken at 20 min and 2 h. Using ultrafiltration (Amicon Centrifree[®] cones), the plasma protein binding of compound **16** to freshly prepared rat and human plasma was determined at 0.2 and 1.0 g/mL. Compound **16** was not highly protein bound, with values ranging from 21–56% in rat plasma and 47–72% in human plasma.

The effects of compound **16** on circadian phase shift were also investigated using the method already described¹⁴ (Fig. 3). Vehicle injection gave no significant phase advance, while melatonin (1 mg/kg) gave a sig-

Table 3. Brain and plasma concentration after IV administration of **16** to rats^a

Time	Plasma (ng/mL)	Brain (ng/mL)	B/P ratio
20 min	5547	2014	0.36
2 h	1002	250	0.3

^aCompound dosed as a solution in PEG-400 at 5 mg/kg, po ($n = 3$).

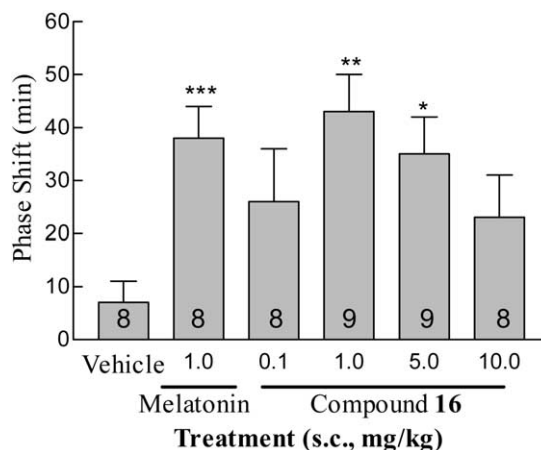


Figure 3. Acute effects of vehicle (5% DMSO, 45% PEG-400, 50% saline), melatonin (1 mg/kg), and **16** (0.1, 1, 5, and 10 mg/kg) on circadian phase shift. Vehicle vs. dose: * $p = 0.005$, ** $p = 0.0003$, *** $p = 0.0002$. The number of rats per treatment is shown within each bar. Rats showing disrupted circadian rhythms were not included in the study. One outlier from the 1.0 mg/kg group and one from the 10 mg/kg were excluded using two standard deviations from the mean.

nificant phase advance of 38 min. Compound **16** at doses from 0.1 to 10 mg/kg produced phase shifts that were not significantly different from that produced by melatonin. The shift in the onset of activity produced by administration of compound **16** at 1.0 and 5.0 mg/kg was significantly different from the small change observed in the vehicle-treated group.

In conclusion, this work has identified novel melatonin agonists with high affinity for MT₁ and MT₂ receptors. Compound **16** is a full agonist that produces circadian phase advances similar to that produced by melatonin. In addition, compound **16** is orally bioavailable and possesses low vasoconstrictive activity in vitro in rat tail artery. Furthermore, compound **16** was chosen as a potential clinic trial candidate.

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References and Notes

- Lerner, A. B.; Case, J. D.; Takahashi, Y.; Lee, T. H.; Mori, W. J. *Am. Chem. Soc.* **1958**, *80*, 2587.
- Yu, H.-S.; Reiter, R. J. *Melatonin: Biosynthesis, Physiological Effects, and Clinic Applications*; CRC Press: Boca Raton, 1993.
- Arendt, J. *Melatonin and the Mammalian Pineal Gland*; Chapman and Hall: London, 1995.
- Arendt, J.; Aldhous, M.; English, J.; Marks, V.; Arendt, J. H.; Marks, M.; Folkard, S. *Ergonomics* **1987**, *30*, 1379.
- Arendt, J.; Aldhous, M. *Ann. Rev. Chronopharmacol.* **1988**, *5*, 53.
- Skene, D. J.; Aldhous, M.; Arendt, J. in 'Sleep '88: Proceedings of the Ninth European Congress on Sleep Research'; Gustar Fischer Verlag, Stuttgart, 1988, p. 39.
- Petrie, K.; Dawson, A. G.; Thompson, L.; Brook, R. *Biol. Psychiat.* **1993**, *33*, 526.
- Maestroni, G. J. M. *J. Pineal Res.* **1993**, *14*, 1.
- Morgan, P. J.; Barrett, P.; Howell, H. E.; Helliwell, R. *Neurochem. Int.* **1994**, *24*, 101.
- Reppert, S. M.; Weaver, D. R.; Godson, C. *Trends Pharmacol. Sci.* **1996**, *17*, 100.
- Takaki, K. S.; Luo, G.; Bertenshaw, S. R. U.S. Patent 6,211,225, 2001.
- The binding assay was carried out following the procedure of Reppert, S. M.; et al. (*Neuron* **1994**, *13*, 1177) with some modification as described below: **Membrane Preparation**: NIH3T3 cells stably expressing melatonin MT₁ or MT₂ receptor cDNA were maintained in DMEM supplemented with 10% calf serum and G-418 (0.5 g/liter). Cells were grown to confluency in T-175 flasks, scraped using Hank's balanced salt solution. Cell pellets were then homogenized in assay buffer (50 mM Tris buffer containing 12.5 mM MgCl₂, and 2 mM EDTA, pH 7.4 at 37 °C with concentrated HCl) with the presence of 10 µg/mL aprotinin and leupeptin, and 100 µM phenylmethylsulfonylfluoride using a dounce homogenizer and centrifuged. The resulting pellet was resuspended with dounce homogenizer in assay buffer (supplemented with the above protease inhibitors), assayed for protein concentration and frozen. On the day of assay, the small aliquot was thawed on ice and resuspended in ice cold assay buffer and held on ice until assayed. **2-[¹²⁵I]-Iodomelatonin Binding Assay**: 2-[¹²⁵I]-Iodomelatonin (0.1 nM and 0.2 nM final concentration for MT₁ and MT₂, respectively), serial dilutions of compound, and membrane preparation were incubated at 37 °C for 1 h. The reaction was terminated by the addition of excess cold wash buffer (20 mM Tris base containing 2 mM MgCl₂, pH 7.4 at room temperature) immediately followed by filtration over glass fiber filters (Whatman). Non-specific binding was defined with 10 µM of cold melatonin. Radioactivity was determined using scintillation counter. The IC₅₀ was defined as the concentration of a compound required to displace 50% of radioligand binding. The values of the equilibrium dissociation constant of the compound (competitor) K_i were calculated from the IC₅₀ values using the equation of Cheng and Prusoff (Cheng, Y.; Prusoff, W. H. *Biochem. Pharmacol.* **1973**, *22*, 3099). $K_i = IC_{50} / (1 + [Ligand] / K_d)$. Data reported in the text are means of 1–4 experiments run at five different concentrations in duplicates. Standard errors were typically within 20% of mean value.
- (a) Viswanathan, M.; Laitinen, J. T.; Saavedra, J. M. *Proc. Natl. Acad. Sci.* **1990**, *87*, 6200. (b) Krause, D. N.; Barrios, V. E.; Duckles, S. P. *Eur. J. Pharmacol.* **1995**, *276*, 207. (c) Ting, K. N.; Dunn, W. R.; Davies, D. J.; Sugden, D.; Delagrange, P.; Guardiola-Lemaitre, B.; Scalbert, E.; Wilson, V. G. *Br. J. Pharmacol.* **1997**, *122*, 1299.
- Mattson, R. J.; Catt, J. D.; Keavy, D.; Sloan, C. P.; Epperson, J.; Gao, Q.; Hodges, D. B.; Iben, L.; Mahle, C. D.; Ryan, E.; Yocca, F. D. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 1199.
- Panlabs SpectrumScreen on more than 80 receptors.